

AMENDMENTS

In the specification:

Please amend the specification as follows:

Please replace Table 2, page 14 with the following.

| Oligo | Sequence ^a | Strand ^b | REN site ^c | Position | SEQ ID NO. |
|------------------|--|---------------------|-----------------------|---------------|------------|
| <i>c-mycl</i> | GATCCATAATGGAGCAAAAGCTCATTT CTGAAGAGGACTTGAATTCTAGATAAC TGCA | + | <i>Xba</i> I | N/A | <u>1</u> |
| <i>c-myc2</i> | GTTATCTAGAAATTCAAGTCCTCTTCAGA AATGAGCTTTTGCTCCATTATG- | - | <i>Xba</i> I | N/A | <u>2</u> |
| CLH1 | CGGCAGATCTATAATGGCGAATGCA | + | <i>Bgl</i> III | 3089- 3100 | <u>3</u> |
| CLH2 | GCCAGGATCCATAATTACAAATAGGAT TGAG | - | <i>Bam</i> HI | 3698- 3718 | <u>4</u> |
| CLH3 | CGGCTCTAGAAATGGCGAATGCATC | + | <i>Xba</i> I | 3089- 3102 | <u>5</u> |
| CLH4 | GCCGCTGCAGTCAATAATTACAAATAG G | - | <i>Pst</i> I | 3704- 3721 | <u>6</u> |
| CLH5 | GGCCGGTACCGAGTTCGTTGACGC | + | <i>Kpn</i> I | 2334- 2347 | <u>7</u> |
| CLH6 | CGCGAGATCTACTTCGCGGCTTCTCGC ACC | - | <i>Bgl</i> III | 3069- 3088 | <u>8</u> |
| CLH7 | GGCCCTGCAGATAATAACAATTATAA AT | + | <i>Pst</i> I | 3722- 3741 | <u>9</u> |
| CLH8 | CGCGAAGCTTAGCAACTATATATT | - | <i>Hind</i> III | 4411- 4424 | <u>10</u> |
| <i>Lef</i> -2PE3 | AAGCTCGTGCCGGAACGCGTGACAG TCG | - | | 2886- 2915 | <u>11</u> |

^a The first first nucleotide corresponding to AcMNPV sequence is ~~shown~~ shown.

^b Relative to the coding strand of *lef*-2.

^c The restriction enzyme site is underlined in the sequence of the oligonucleotide.

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|-----------|--------------------------------------|---|-------|---------------|-----------|
| Lef-2PE4 | TGTAGTCGGCAGTTCATTTTGGGCGTG ATCG | - | | 2966- 2995 | <u>12</u> |
| Lef-2RT1 | AAGAAAACAATGTACCGCGCGGCGG | + | | 3438- 3462 | <u>13</u> |
| Lef-2stop | ATGCGAATTCTCAATAATTACAAATAG GATTG | - | EcoRI | 3700- 3721 | <u>14</u> |

Please replace Table 3 on page 19 with the following.

Table 3: Synthetic oligonucleotides used to construct recombinant viruses

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| Oligo | Sequence (5'-3') | SEQ ID NO. |
|--------|--|---------------|
| SwaIF | AAATTCAGATATAAAGACGCTGAAAATCATTG | <u>15</u> |
| SwaIR | TGATTTTCAGCGTCTTTATATCTGAATT | <u>16</u> |
| AvrIIF | ATTTTCGCTCTAACATAACCACCCTAGGGATGTAC | <u>17</u> |
| AvrIIR | GTACATCCCTAGGGTGGTATGTTAGAGCGAAAATCAAA | <u>18</u> |
| SnaBIF | CTAGGGATTATAAATTTAATGAATTATTAATAATAC | <u>19</u> |
| SnaBIR | GTATTTTAATAATTCATTAAATTTATAATCC | <u>20</u> |

Please replace paragraph 2 on page 22 with the following.

B3

The *AcΔ.lef-2.sup4-o⁹* was used to derive a recombinant virus with a human C-MYC coding region added to that of *lef-2*, to permit the identification of LEF-2 in virus-infected cells. The C-MYC epitope or tag comprises 12 amino acids (SEQ ID NO: 26 NH₂-MEQKLIEDLNSR-COOH), which can be recognised by a monoclonal antibody (9E10; Evan *et al.*, 1986). It was not known whether addition of this epitope would affect the folding and/or the activity of LEF-2 when added to either end of the protein.

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Two transfer vectors were constructed containing the *c-myc* coding sequence at either the 5' (p*Aclef-2.c-myc5'*) or 3' (p*Aclef-2.c-myc3'*) ends of the *lef-2* coding region (Fig. 2). These were used individually to co-transfect insect cells with *AcΔ.lef-2.sup4-o⁹* DNA isolated from yeast cells. After 6 days, the cell culture medium for each sample was titrated in a plaque assay to monitor virus production. Infectious virus (*Aclef-2.5'c-myc*) was only produced by cells co-transfected with *AcΔ.lef-2.sup4-o⁹* and p*Aclef-2.c-myc5'*. This was amplified further in insect cells to derive a virus stock (*Aclef-2. c-myc5'*) of comparable titer to that produced by AcMNPV (ca. $1-2 \times 10^8$ p.f.u./ml)

Please replace paragraph 2 on page 23 with the following.

B4

The transcription start sites for *lef-2* in AcMNPV-infected cells were determined by primer extension using two different oligonucleotides. Primer *lef-2PE4* was used to detect the early start site. An extension product was obtained with mRNA extracted at 6 and 12 hr p.i., corresponding to a start site located 279 bp upstream from the *lef-2* ATG codon. This mapped at a C within the sequence SEQ ID NO: 21 CAATGCGCCCGTTGT (Fig. 4A). When virus-infected cells were treated with cycloheximide, no products were obtained in the subsequent primer extension analysis, underlying the early character of this promoter. Primer *lef-2PE3* was used to detect the late start site. One major extension product was obtained with mRNA extracted at 12 and 24 hr p.i. (Fig. 4B). This mapped the transcription start site at an adenine within a TAAG motif located 361 bp upstream of the ATG. When mRNA was extracted from virus-infected cells treated with aphidicolin, no primer extension products were obtained. Identical results were obtained with mRNA from *Aclef-2.c-myc5'*-infected cells (data not shown).

Please replace paragraph 3 on page 31 with the following.

B5

Gene expression was compared in cells infected with AcMNPV or *Aclef-2c-myc5'*, to determine if the addition of the tag to LEF-2 had any effect on transcription. Transcription analysis of *lef-2* in AcMNPV and *Aclef-2c-myc5'*-infected cells showed no differences between the two viruses. In both viruses, a dual promoter included within the

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350 nucleotides upstream of the ATG drove the expression of *lef-2*. The *lef-2* transcripts were detected as early as 1 hr p.i. and reached a high level by 48 hr p.i. The early start site was mapped at a C within the sequence SEQ ID NO: 21 CAATGCGCCCGTTGT localised 279 nt upstream of

